

# Early biomarkers indicate COPD induced by whole cigarette smoke in live human lung tissue

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## Introduction

Cigarette smoke (Cs) inhalation is a main reason to develop chronic obstructive pulmonary disease (COPD). It is characterised by degradation of alveoli, inflammation and mucus hypersecretion. Mechanisms that underlie various components of COPD can be modelled *in vitro*, specifically using cigarette smoke with fresh human lung tissue. The aim of the study is to establish pathological changes of COPD in vital lung tissue by using Cs and cigarette smoke condensate (Csc).

## Materials and Methods

Human Precision-Cut Lung Slices (PCLS) were exposed to Csc or whole Cs in an Air-Liquid Interface (ALI) using the *in vitro* exposure device P.R.I.T.® ExpoCube®. Cytotoxicity, release of cytokines and extracellular matrix (ECM) proteins were analysed. Pharmacological treatments were applied to inhibit inflammatory responses of tissue to Cs.

## Results

Concentration dependent cytotoxicity was observed in human PCLS after 24 h Csc submerge exposure and cigarette smoke exposure in ALI culture. EC<sub>50</sub> values were determined using WST-1 assay. EC<sub>50</sub> values of 196 µg/mL for Csc and 16 µg particles deposited on human lung tissue for Cs were calculated (Fig. 2).

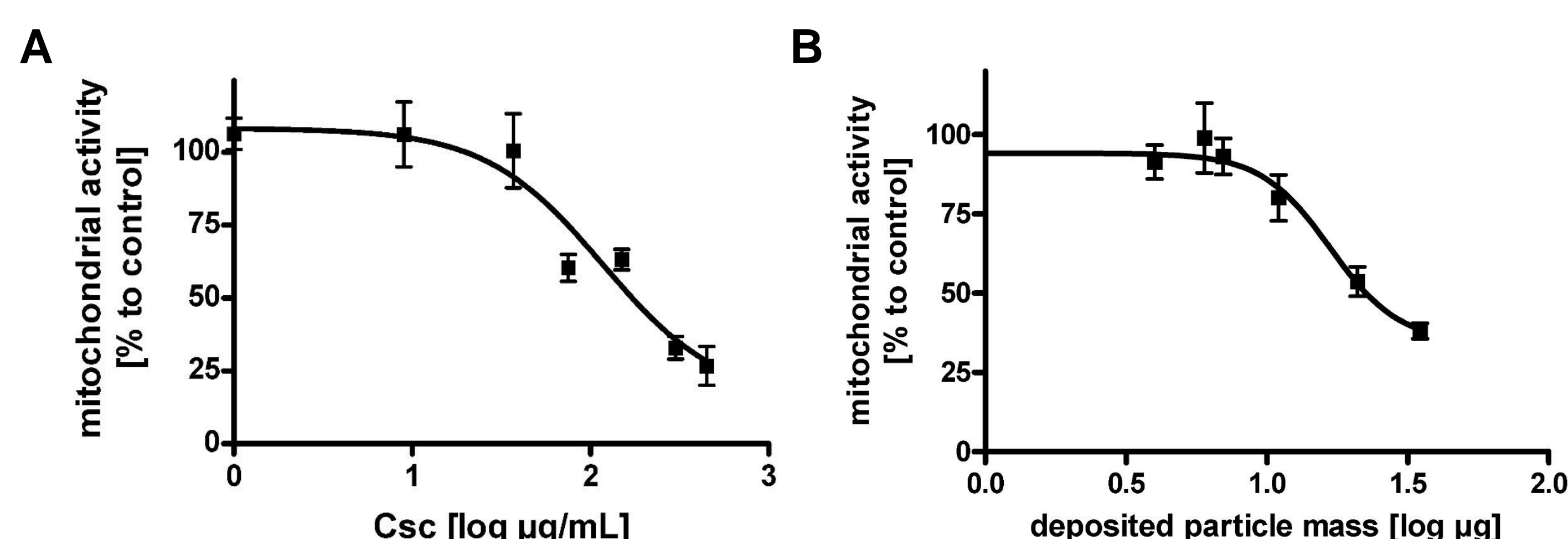


Fig. 2: Viability of human PCLS after 24 h exposure to increasing concentrations of Csc (A) or Cs (B) was assessed by WST-1 assay. N=4 for Csc, N=3 for Cs, technical duplicates.

Human lung tissue exposed to clean air (0 µg) shows no loss of viability, as did the respective non-exposed lung tissue control. PCLS exposed to the smoke of two cigarettes (7 µg) shows loss of viable tissue and an increase in dead cells (Fig. 4).

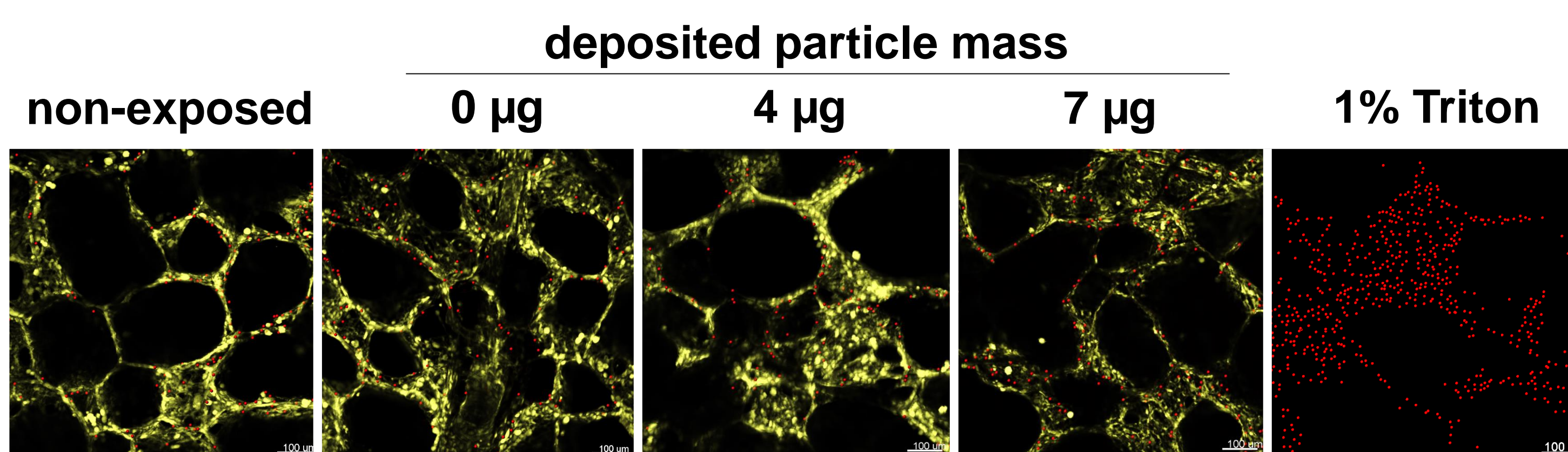


Fig. 3: Viability of lung tissue exposed to increasing concentrations of Cs was assessed by LIVE/DEAD® staining. Viable lung tissue stained with calcein was detected in non-exposed control (yellow). Dead cells were stained using ethidium homodimer-1 (red), shown in Triton control.

Pro-inflammatory cytokines interleukin (IL-) 1α and 1β and matrix metalloproteinases (MMP-9) were analysed by ELISA (Fig. 4). Significant increase of pro-inflammatory cytokines can be observed after 24 h Csc exposure of lung tissue. Dexamethasone inhibited the Csc-induced production of IL-1α.

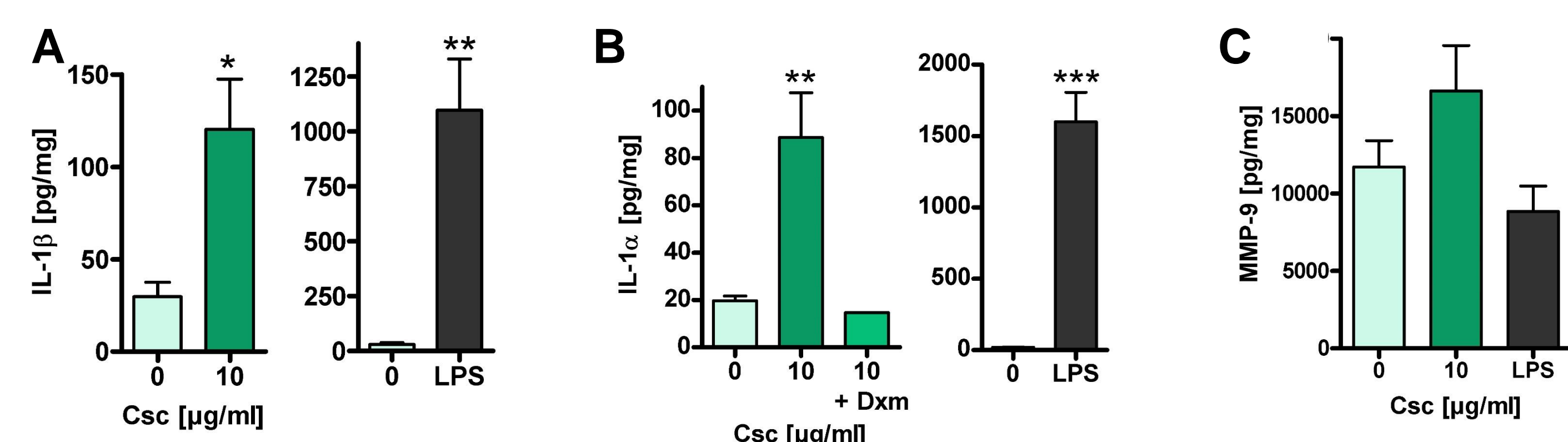


Fig. 4: Production of cytokines in PCLS after Csc exposure or Csc + Dxm was quantified by ELISA. Statistical significance is indicated by \*p<0.05, \*\*p<0.01, \*\*\*p<0.001. N=3.

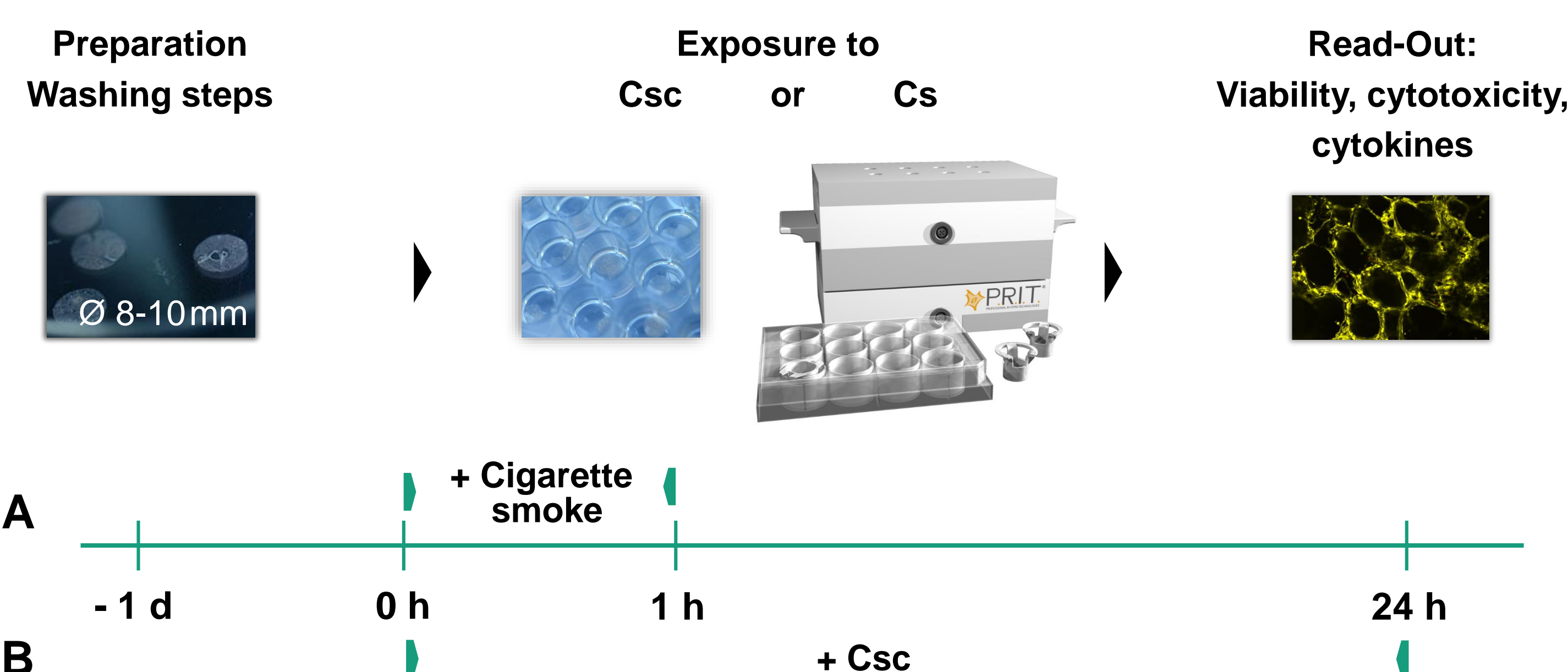


Fig. 1: Experimental setup for submerge exposure to Csc for 24 h (B) and for exposure of lung tissue to cigarette smoke at ALI (A).

Exposure of human lung tissue to Cs significantly increased the release of pro-inflammatory cytokines (Fig. 5). MMP-9, Pro-Col1α1 and extracellular RAGE present significant changes in the ECM after Cs exposure (Fig. 6A-C). Increased ratio of MMP-9 to TIMP-1 are biomarkers for a emphysema development (Fig. 6D). Pharmacological intervention reduced the Cs-induced inflammatory cytokines. Dexamethasone significantly reduced IL-1α production (Fig. 6E, F).

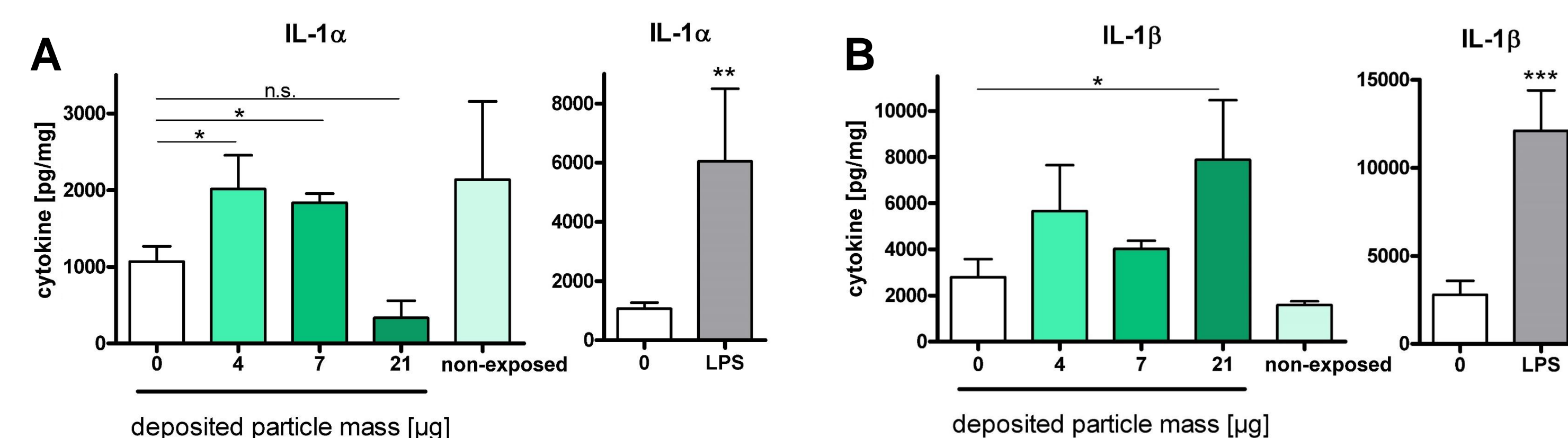


Fig. 5: Production of pro-inflammatory cytokines in PCLS after Cs exposure were quantified by ELISA. Statistical significance is indicated by \*p<0.05, \*\*p<0.01, \*\*\*p<0.001. N=3.

